FAQ SHEET



PERSONAL PRECISION

What is the minimum hold time for thermal cycling?	For holds during thermal cycling, the minimum hold time is 3 seconds.
vendors?	Yes. However, in order to ensure optimal performance, please ensure that any 3rd party disposables:
	1) Are mechanically identical to the standard MyGo disposables, to ensure a good fit in the mounts and correct operation of the heated lid.
	2) Provide vapour tight seals when closed.
	3) Are optically transparent.
	4) Are biocompatible, and free of DNAse and RNAse contamination.
How do I create a template?	All experiments can be used as a template. Click "Open", then use the "file type" selection in the file dialog to select the "Template" option, then select the file to open as a template.
What quenchers can I use in the instrument?	For best results use non fluorescent quenchers, for example BHQ.
Can I clean the wells to remove potential contaminants?	Please contact technical support.
Do I need to optically calibrate the instrument before using it?	The MyGo Mini comes pre-calibrated for 11 dyes. If the dyes you want to install are pre-calibrated then you do not have to run a dye calibration.
When should I generate a dye file?	We would recommend generating dye files if you are using a dye that is not present on the list of pre-calibrated dyes. In addition, generating dye files can help with multiplexing or other challenging applications.
Can I use dye files across multiple instruments?	Yes, dye files can be used on different instruments of the same model (i.e. MyGo Mini S).

Why do some amplification curves disappear when I change from Full to Background normalisation in Auto Quant?	In Full normalisation mode the scale of low intensity amplifications is boosted. In Background normalization mode the scale of low intensity amplifications is not boosted.
Can my dye files be used on historical runs?	Yes. To add new dye files to existing experiments, go to the "Run Settings" tab and select the "Display Dye Calibrations" checkbox, then click the "Open" button to import a dye calibration file.
How will I know if there is a new software version?	The latest version of software is available at www.mygopcr.com
Do the MyGo instruments conform to the electrical safety requirements of IEC 61010 or equivalent and the electromagnetic compatibility requirements of IEC 61326 or equivalent?	Yes
Will my instrument run with empty tubes?	Yes. The MyGo Mini does not need PCR reagents to be present before performing a run.
I only want to run one tube in my MyGo Mini. Is this possible?	Yes, please ensure that tubes, with or without reactions are present in wells 1, 5, 9 and 13.
Can I change the run settings once a run has started?	No. Once the run has been started, optical and thermal settings can not be changed.
Can I start a run before setting up my samples and targets information?	Yes. You can start an experiment without setting up samples and targets.
Can I change how my samples and targets are set up while my experiment is running?	Yes, the MyGo Mini will acquire data from all 3 optical channels in every well at every acquisition. This means the user can view raw data from all channels in all wells. You are not restricted to data from a channel chosen at the start of a run.
Do the instruments have built in memory?	Yes. This memory provides for a data buffer during runs to reduce the risk that data is lost due to a poor connection between your PC and the instrument. Please ensure that you save your data on your PC once your run has finished.
Why don't the MyGo instruments have power switches?	All moving parts, including power switches, represent potential points of failure for instruments. For this reason MyGo instruments do not have power switches. MyGo instruments are designed to be left connected to a power supply at all times. When not in use MyGo instruments enter a low power standby mode. If you need a power switch for your MyGo instrument, then they are available as accessories.

Do I need to use ROX as a passive reference in my MyGo instrument?	Some real-time PCR instruments require the use of ROX as a fluorescent passive reference to correct for optical artefacts. Generally, modern instruments do not require the use of passive fluorescent references. The MyGo instruments do not require the use of ROX as a passive reference.
My LAN is unreliable, how can I reduce the risk of losing data?	You can run your instrument directly from a USB drive, to which data will be saved.
Where is the threshold for determining Cq values in Auto Quant?	Modern methods of determining Cq values are not based on simple thresholds. Modern methods of Cq determination are model based. Auto Quant fits a model of a PCR amplification to the fluorescence data observed. This model fit then enables the estimation of a number of important parameters including Cq values.
Why do some of my amplification curves drop down before rising in Auto Quant?	Correct background removal in Auto Quant relies on correct qualitative calling. Please check that appropriate thresholds are set so that positive amplifications are called positive.
Is it normal for the lid of my instrument to get warm during use?	Yes.
Can I run the MyGo Mini from a battery?	Yes. A power supply unit, that enables the MyGo Mini to run from a 12V automotive battery, is available as an accessory.
How many MyGo Mini instruments can I control from one PC?	You can control up to 400 MyGo Mini instruments from one PC.
What does integration time mean?	"Integration time" is the exposure time used by the camera in your MyGo instrument when measuring fluorescence.

What kind of computer is required to run the MyGo Mini software?

The MyGo software runs on windows, Mac and Linux and does not require a dedicated PC or laptop.

PC Requirements

The table below outlines minimum computing requirements to run the MyGo software.

Processor	Intel Core i3 2GHz (or equivalent AMD)
Memory	8 GB
Hard Disk	250 GB
LAN	RJ-45 LAN (100MBit)
USB	USB 2.0 (if using USB for runs)
Display Resolution	Display Resolution 1280* 1024 of greater
Operating System	Microsoft Windows 8 or 10

